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- 1. Open a blank Powerpoint presentation; and click on "insert", then on "Text Box". Type in the title and authors and move the text box to the top and center.
 - 2. Next click on "insert" then on "Picture" and insert your logo or the logos of any of the other authors
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ABSTRACT

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The great blue heron (GBH; Figure 1) population is stable in the Chesapeake Bay. These majestic colonial water birds are symbolic of wetland and Bay health and a welcome sign to visitors enjoying the Eastern Shore of Maryland. On October 5, 2001, the first GBH was collected from Poplar Island (Figure 2). Throughout the next 2 months, 2 birds were found in Anne Arundel County, and 6 in Queen Anne's County (Figure 2). Most live birds were taken to the TSBRR facility in Newark, DE. Birds presented with clinical signs of emaciation, lethargy, inability to fly, and an unusually hard abdomen. Upon admission to the rehabilitation facility the blood profiles (Table 1) revealed anemia and varying degrees of hypoproteinemia, and all birds were dehydrated and depressed, were in lateral recumbency, and had profuse diarrhea. At least two birds were in respiratory distress. Due to poor prognosis, euthanasia was performed and necropsies were conducted on all birds (Table 2).

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Steatitis is defined as an inflammation of adipose tissue 1. In mammals, it is associated with a deficiency of Vitamin E and /or Selenium. Vitamin E and selenium have antioxidant properties and are essential to cell membrane integrity². Interaction between Vitamin E / Selenium and dietary unsaturated lipids likely play a role in the pathogenesis of steatitis though the exact mechanism is unknown. In birds, this condition is suspected to be caused by a diet high in rancid or oily fish containing polyunsaturated fats3. Steatitis has been reported in several other bird species (black crowned night heron Nycticorax nycticorax, osprey Pandion Haliaetus, double crested cormorant Phalocrocorax auritus), 3,4,5,6

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While analysis of all bird tissues was the goal, due to monetary constraints, only the tissues from those birds dying in the GBH event were tested for microcystin. Liver samples from six of the eight GBH in custody were sent to Wright State University for cyanobacterial toxin identification using immunoassay (ELISA), plus liquid chromatography/mass spectrophotometry (LC/MS). Results of the ELISA and LC/MS toxin analyses detected microcystin in 5 of the 6 samples. Toxic levels of microcystin known to cause acute lethal toxicosis were found in 4 of the 6 tissue samples (Table 1). The significance of these findings is currently under investigation. Monitoring of bird populations in this geographic area is ongoing and will be conducted over the next few years, depending on the level of funding available.

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NOTE: this poster is a little too wordy and has more references and acknowledgments than are usually needed.





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Table 1. Blood/tissue chemistry and Body Condition Index (BCI) for 6 great blue herons collected from the Chesapeake Bay during the Fall 2001

ID#	SEX	Location	Wgt (g)	HCT/BC/ TS *	BCI **	Heavy Metals	O.C	Blood Chemistry	Microcystin ^A (ng/g)
WHAHE20	N/A	Poplar Is.	1270	23/2/3.0	2	(lead only) Normal	Neg	Normal	BDL ^B
WHAHE22	М	Centerville	2270	N/A	3	Normal	Neg	Normal	4.85
WHAHE16	М	Carr's Crk.	2245	12/1+/3.2	3	N/A	N/A	N/A	1.65
WIAHE25	М	Carr's Crk.	2905	18/1+/1.6	4	(Zn & Fe) High	Neg	Normal	1.43
WHAHE24	М	Stevensville	2960	20/2+/3.0	4	Normal	Neg	Normal	1.25
WHAHE27	м	Stevensville	2610	N/A	3	N/A	N/A	N/A	0.95

- *HCT = Hematocrit (% red blood cells)
- BC = Buffy coat (% white blood cells TS = Total solids expressed in g/dL
- **BCI = Body Condition Index, scale of 1-5 (1 = emaciated, 3 = normal, 5 = fat).
- ***Organochlorine =GC/MS/GC screens that detect insecticides (organophosphates, carbamates and organochlorines), strychnine, metaldehyde, several drugs, environmental contaminants.

Table 2. Case summaries (5 GBHs presented to TSBRR)

	Presentation	
•Not standing (5/5)	•Firm, "ropey" abdomen (5/5)	• Open wound on back (1/2
•Yellow-brown diarrhea (5/5)	•Agonal respiration (1/5)	• Covered in mud (1/5)
	Treatment on entry	
•Dexamethasone 3mg/kg (1/3)	•Hetastarch PO (2/	3)
•2.5% Dextrose in Lactated Rin	ger's Solution, 35-40cc IV (3/3)	

•Ivermectin 0.2mg/kg SQ once (1/3)	•Metronidazole 50mg/kg PO BID (3/3)
•Vit E 200IU PO BID (2/3)	•Selenium 200mcg PO SID (1/3)

•Gavage feed prn (3-5% of body weight, up to 3x/day) if not self-feeding on fish (3/3)

Time in care (avg.. = 3 days; range = 0-7 days)

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Figure 2. Location of avian mortality events, harmful algal blooms, and sites where debilitated great blue heron were collected in the Chesapeake Bay during the Fall 2001.

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Figure 3. External view of GBH abdomen with fat deposits visible through the skin.

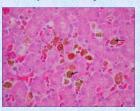


Figure 5. Microscopic view of extensive fat cells



Figure 4. View of GBH body cavity showing extensive subcutaneous and abdominal fat deposits.



Figure 6. Cyanobacterial bloom where several

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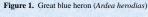
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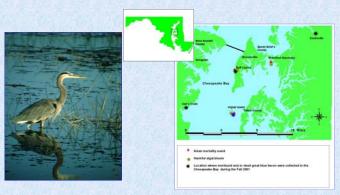


Figure 1. Great blue heron (Ardea herodias)

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Figure 3. External view of GBH abdomen with fat deposits visible through the skin



Figure 4. View of GBH body cavity showing extensive subcutaneous and abdominal fat deposits.

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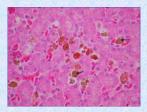


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•Hetastarch PO (2/3) •Dexamethasone 3mg/kg (1/3)

•2.5% Dextrose in Lactated Ringer's Solution, 35-40cc IV (3/3)

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Time in care (avg., = 3 days; range = 0-7 days)

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INVESTIGATION

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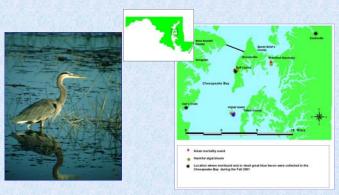


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Microcystins are cyanobacterial hepatotoxins found in a number of cyanobacterial genera, are widespread worldwide, favor inland waters and nutrient-rich, warm brackish waters.

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On October 18, 2001 a GBH was euthanized and sent to the National Wildlife Health Center, Madison, WI, for a post-mortem examination. Remaining birds were necropsied at the MD DNR Cooperative Oxford Laboratory or at TSBRR. Consistent necropsy findings included emaciation, decreased muscle mass, pale muscle color, fat atrophy, gastro-intestinal parasitism, and excessive deposits of waxy yellow fat in the abdomen, subcutis, and throughout the body cavity (Figures 3 and 4). A gross necropsy diagnosis of steatitis was determined to be the consistent finding in all carcasses. Selected tissues were sent to the Armed Forces Institute of Pathology for histopathological confirmation of steatitis (Figure 5).



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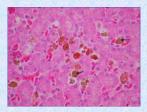


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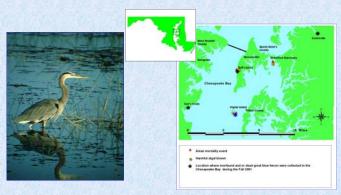


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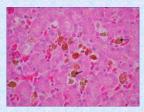


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