In a large animal practice, it seems that I spend most of my time trying to get the right ova with the right sperm in the right uterine environment at the right time of the year in order to get the right neonate born in the right environment with the right provisions to get the right adult so I can do it again. Can that be wrong? I think it’s only right! Well, that is kind of what Dr. Gary Althouse and the symposium chairs have tried to do for the Annual Conference in Colorado Springs. The idea was to put the right information together from the right speakers for the right people in the right location at the right time of year, and I am excited, because they have gotten it right. Time draws closer to the Annual Conference in beautiful Colorado Springs. All that knowledge and information from the conference, combined with the natural beauty of Colorado, makes for a rare opportunity that you cannot afford to miss. Since we were unable to gather in Vancouver, this year’s conference should be first on everyone’s list of CE and social event calendars. Be real sure it is at the top of your list.

The conference will be preceded with board meetings where your directors will continue to focus on the future of the SFT. During my time on the board of directors, the goal has been to bring the SFT forward into the future with a definitive plan in finance and service. We have not been able to plan for all the things that have happened, but a strong effort has been made to continue on with the resolution that the SFT will flourish through any difficulties while still providing better service to its membership. Gary Althouse and Fred Lehman are already discussing future planning strategies and methods. With the board of directors and officer’s hard work, the future of the SFT looks extremely bright. They now need the input of the membership. Some great ideas and suggestions have been brought forward by the membership this year. Thank you for those, but don’t stop now. We need your input now more than ever. Please contact one of the board members or me with any suggestions you have before the beginning of the Annual Conference. Help us better serve you.

Student membership is really taking off. I have never seen the interest and enthusiasm that these students have for Theriogenology. It looks like we will have a good student attendance at Colorado Springs, and I couldn’t be more pleased. These young professionals are the future of the SFT, and their presence will bring a new energy to this stand-alone meeting.

The stage is set for one of the most enjoyable and information packed Annual Conferences yet. Come spend five wonderful Theriogenology packed days with us in Colorado Springs. Look over the program (online at www.therio.org), make your reservations, and join us for a great time with some great people.
Over these last few months Walker Management, session chairs, symposia chairs and I have been actively working to finalize arrangements for the upcoming 2002 SFT Annual Conference. The 2002 locale is at the Adam’s Mark Hotel in Colorado Springs, CO, with the meeting convening from August 7-11th. You can now get complete details on the upcoming symposia and conference, including a list of the speakers and their topics, at the SFT website (www.therio.org). Along with complete information on the meeting, you can also register on-line at this site. Early registration is available right now and extends through July 8, 2002 for all symposia and the annual conference. And, as an added benefit for this year’s meeting, the conference proceedings will be available on CD-ROM. So don’t delay, go to www.therio.org and sign up for what is sure to be an exciting and informative meeting. I look forward to seeing you there!

SFT would like to thank those members who made contributions to the Society after the necessary cancellation of the Annual Conference last September. The 2002 membership dues statements had an option for making a gift to SFT, and many members extended their generosity by making a contribution. Through April of this year, more than $2,000 has been contributed to the Society, which will assist in supporting membership programs. The SFT Board of Directors and staff would like to thank all individuals who have helped.
AUGUST 7-11, 2002 • ANTLERS ADAM’S MARK HOTEL • COLORADO SPRINGS, COLORADO
2002 SFT ANNUAL CONFERENCE & THE SFT/ACT SYMPOSIA REGISTRATION FORM

Please copy this form for additional registrants. Registration fees include proceedings and scheduled breaks.

Name ___________________________________________________________________________________________________________

Business/Clinic Name __________________________________________________________________________________________________

Street Address __________________________ P. O. Box _____________ City __________ State ______ Zip/Postal Code_____________Country ______

Phone __________________________________ Fax __________________________ Email _______________________________________

(Required for confirmation purposes.)

Please mark here if this is new contact information which needs to be updated in the SFT database. Conference attendees will be posted on the SFT website. If you do NOT want your name listed, mark here.

I. 2002 SYMPOSIAS

With the exception of Canine, Symposia fees do not include admission to the Annual Conference:

By July 8 After July 8

• Bovine BSE Symposium
  Members $200 $270 $ _____
  Non-Members $285 $355 $ _____

• Technician/Breeder Canine Symposium $50 $50 $ _____

• Canine Symposium (Price includes registration to the Annual Conference)
  Members:
    Lecture Only $200 $270 $ _____
    Lecture & Wetlab* $460 $530 $ _____
  Non-Members:
    Lecture Only $285 $355 $ _____
    Lecture & Wetlab* $545 $615 $ _____

*Space in the wetlab is limited. If your form is received after the lab is full, you will be notified and given the option to cancel or to be placed on a waiting list.

• Toxicology Symposium
  Members $300 $375 $ _____
  Non-Members $385 $455 $ _____

II. ANNUAL CONFERENCE

By July 8 After July 8

Members $200 $270 $ _____
Non-Members ** $285 $355 $ _____

** Includes a one-year SFT membership for veterinarians, please check here only if you wish to decline the membership offer (declining membership does not reduce your registration fee).

TOTAL AMOUNT DUE $ _____

PROCEEDINGS FORMAT

Please indicate below, the format in which you would like to receive your proceedings.

(If you do not mark a preference, you will automatically receive a CD-ROM)

❏ Bound Book Version ❏ CD-ROM

PAYMENT INFORMATION

You may register via fax (615/254-7047) or SFT website (www.therio.org) if you are paying by Visa or Mastercard. Otherwise, please mail your payment and completed registration form to: SFT Headquarters, 200 Fourth Avenue North, Ste 900, Nashville, TN 37219.

Fees are due and payable to SFT only in U.S. Funds drawn on U.S. Banks. Please indicate your method of payment:

❏ Check ❏ Visa ❏ Mastercard

Card Number _______________________________________________________________________
Expiration Date ___________________________________________________________________
Cardholder’s Name __________________________________________________________________
Cardholder’s Signature __________________________________________________________________

SPECIAL NEEDS/DIETARY REQUIREMENTS

If you require any special assistance or auxiliary aids to attend this meeting please contact the SFT Office no later than July 8, 2002. Should you have any special dietary requirements, please indicate below:

❏ Vegetarian ❏ Vegan ❏ Fruit ❏ Food Allergies — please specify ______

For Office Use Only

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If you have questions, please contact SFT Headquarters at 615/244-3060.
Or e-mail us at sft@walkermgt.com
Guide Dogs for the Blind’s Director of Canine Health and Training, Patricia Olson, DVM, PhD, has been named by the Society for Theriogenology (SFT) Board of Directors as the recipient of the 2002 Bartlett Award. She is recognized for outstanding service to the reproductive veterinary community, as well as her years of support for the SFT.

As the Bartlett Award winner, Dr. Olson will be recognized at the 2002 SFT/ACT Annual Conference and Symposium, scheduled for August 7-11 in Colorado Springs, CO. She will also present at the meeting, which will take place Friday, August 9 at 7:00 pm, with a reception immediately following.

Dr. Olson was born and educated in Minnesota, receiving B.S., D.V.M. and M.S. degrees at the University of Minnesota, Twin Cities Campus. She received a PhD in reproductive physiology and endocrinology at Colorado State University and was president of the American College of Theriogenologists from 1993-94. From 1995-1998, she was the Director of Veterinary Affairs and Studies for the American Humane Association. She recently co-authored a new textbook (published by WB Saunders, Co.) Canine and Feline Theriogenology, which addresses the reproductive health of dogs and cats, and why every puppy and kitten born should have the best health, the best care, and the best opportunity for a good life.

In 1993-1994, Dr. Olson worked at the United States Senate as an AVMA Congressional Science Fellow. During her senate work, she investigated potential causes of Gulf War Syndrome, and designed a study for evaluating the effect of reproductive toxicants on the many men and women who served in the military. The results of this study led to recommendations on reducing exposure to substances that could result in birth defects for veterans’ children.

Earlier this year, Dr. Olson accepted the new position of Director of Canine Health and Training with Guide Dogs for the Blind, Inc. The national nonprofit provides skilled Labrador Retrievers, Golden Retrievers, German Shepherds, Lab/Golden Crosses, and training in their use free of charge to blind and severely visually impaired people. It is one of the largest schools of its kind in North America.

Please make plans to attend the SFT Annual Conference, and hear Dr. Olson present as the 2002 Bartlett Award recipient.

Make sure to mark your calendar, for SFT’s Future Meetings

September 16-20, 2003
SFT Annual Conference & Symposium
Columbus Convention Center, Columbus, OH

August 4-8, 2004
SFT Annual Conference & Symposium
Hyatt Hotel, Lexington, KY
The Auburn SFT Chapter recently hosted the Auburn Hudson-Walker Theriogenology Conference. The Conference was a tremendous success, and resulted in a significant increase in Theriogenology interest. In fact, over 70 Auburn students joined SFT!

Auburn SFT Student President, Lisa Willis (right) and the chapter’s Vice President, Justin Murray

Theriogenology Conference Speakers

Call for Articles/Stories From Students – Let Us Hear From You!

If you have news from your SFT Student Chapter that you would like to share with SFT members, please send it to sft@walkermgt.com. Please make sure to put "SFT Newsletter Article" in the "subject" line of your message.

Also, please keep in mind that the Society for Theriogenology (SFT) provides student chapter grants. The funding is intended to provide educational advancement of students interested in Theriogenology. Furthermore, primary consideration will be given to funding requests which provide hands-on educational experience for students. The SFT may consider all or partial funding of a request based on the annual budget and the merit of the educational experience to students.

The request for funding student grants:
• May come from the SFT appointed faculty advisor and/or student chapter members of the SFT
• A faculty member should sign the request or send a letter of support
• Must be for the sole benefit of students in a Professional Veterinary training program
• Must be specific to the study of Theriogenology
• Should include a cover letter, a course description and a budget

For more information, please contact the SFT office at sft@walkermgt.com or 615/244-3060.
SFT Recognizes Two New Honor Roll Members

The Society has recently inducted its 20th and 21st Honor Roll members, Dr. John Bowen of Blacksburg, VA and Dr. Lawrence Rice of San Antonio, TX.

Congratulations to Drs. Bowen and Rice!

SFT Announces Board Candidates

SFT is pleased to announce the Board of Directors candidates as forwarded by the Nominations Committee. The individuals below are running to serve a three-year term as a director on the SFT Board of Directors. Pursuant to the SFT Bylaws, the general election will be held during the Annual Business Meeting at the next Annual Conference (August 7-11 in Colorado Springs).

Dr. Carlos Risco – University of Florida, Gainesville, Florida
Dr. Tom Riddle – Rood & Riddle Equine Hospital, Lexington, Kentucky
Dr. Ahmed Tibary – Washington State University, Pullman, Washington

Completing their term on the Board this August are Drs. Nikola Parker, John Shull and Dirk Vanderwall. SFT would like to thank these individuals in advance for their dedication and service as directors.

Follicle Selection in Cattle: Dynamics of Follicular Fluid Factors During Development of Follicle Dominance

Follicle diameter deviation during follicular waves in cattle begins with a reduction in growth rates of developing subordinate follicles, in contrast to the maintenance of a constant growth rate by a developing dominant follicle. In experiment 1, the temporal changes encompassing deviation in concentrations of follicular fluid factors relative to one another in the three largest follicles (F1, F2, and F3) were studied. Follicular fluid samples were collected when F1 reached diameter ranges of 7.0-7.9, 8.0-8.9, 9.0-9.9, and 10.0-10.9 mm (n = 12 per range). The first increase (P < 0.05) in the difference between F1 and F2 for estradiol occurred at the 8.0- to 8.9-mm range, which was one range earlier than for diameter (P < 0.05). Free insulin-like growth factor (IGF)-1 concentrations in F1 were similar among diameter ranges, but concentrations in F1 were higher (P < 0.05) than in F2 for each range except 7.0-7.9 mm. Concentrations of free IGF-1 in F2 decreased (P < 0.05). No significant differences were detected in concentrations of progesterone, androstenedione, total inhibin, and inhibin-A. Averaged over follicles, inhibin-B decreased (P < 0.05) between the 8.0- to 8.9- and 10.0- to 10.9-mm ranges, and activin-A increased (P < 0.05) between the 7.0- to 7.9- and 9.0- to 9.9-mm ranges. However, no differences were found among follicles. In experiment 2, changes associated with the development of dominance by F2 were studied using ablation of F1 at the beginning of expected deviation (F1, 8.5 mm; Hour 0) as the reference point. Follicular fluid factors were compared at Hour 12 between F2 of a control group (F1 intact; n = 10) and an ablated group (F1 ablated; n = 10). Diameter (P < 0.02), estradiol (P < 0.001), free IGF-1 (P < 0.002), and progesterone (P < 0.003) were greater and IGF-binding protein-2 was lower (P < 0.01) in F2 of the ablated group at Hour 12. No differences were detected in concentrations of androstenedione, total inhibin, and inhibin-A. The results of the two experiments indicated, on a temporal basis, that intrafollicular changes in estradiol and the IGF system, but not in the inhibin/activin system, could account for a reported greater FSH responsiveness by the future dominant follicle than by the future subordinate follicles by the beginning of diameter deviation in cattle.


DISCLAIMER

The Society for Theriogenology does not take responsibility for information contained in or accuracy of the Abstracts published in this newsletter.
Reproductive Characteristics of Cloned Heifers Derived from Adult Somatic Cells

Enright, BP; Taneja, M; Schreiber, D; Riesen, J; Tian, XC; Fortune, JE; Yang, X (2002): Biol. Reprod. 66, 291-296.

This study examined the onset of puberty, follicular dynamics, reproductive hormone profiles, and ability to maintain pregnancy in cloned heifers produced by somatic cell nuclear transfer. Four adult somatic cell-cloned heifers, derived from a 13-yr-old Holstein cow, were compared to 4 individual age- and weight-matched heifers produced by artificial insemination. From 7 to 9 mo of age, jugular venous blood samples were collected twice weekly, and from 10 to 11 or 12 mo of age, blood sampling was carried out every other day. After the heifers reached puberty (defined as the first of 3 consecutive blood samples with peripheral plasma progesterone concentrations of >1 ng/ml), ultrasound examination of ovaries and jugular plasma sample collection were carried out daily for 1 estrous cycle. Cloned heifers reached puberty later than controls (mean +/-SEM, 314.7 +/- 9.6 vs. 272 +/- 4.4 days and 336.7 +/- 13 vs. 302.8 +/-4.5 kg for clones and controls, respectively; P < 0.05). However, cloned and control heifers were not different in estrous cycle length, ovulatory follicle diameter, number of follicular waves, or profiles of hormonal changes (LH, FSH, estradiol, and progesterone). Three of the 4 clones and all 4 control heifers became pregnant after AI. These results demonstrate that clones from an aged adult have normal reproductive development.

Responses of Seasonally Anovulatory Mares to Daily Administration of Thyrotropin-releasing Hormone and (or) Gonadotropin-releasing Hormone Analog.


Seventeen seasonally anovulatory light horse mares were treated daily, starting January 5 (d 1), for 28 d with GnRH analog (GnRH-A; 50 ng/kg BW) and(or) thyrotropin-releasing hormone (TRH; 5 ug/kg BW) in a 2 x 2 factorial arrangement of treatments to test the hypothesis that combined treatment may stimulate follicular growth and development. Ovaries were examined via ultrasonography and jugular blood samples were collected every 3 d. Frequent blood samples were collected after treatment injections on d 1, 2, 4, 7, 11, 16, and 22; on d 29, all mares received an i.v. mixture of GnRH, TRH, sulpiride, and EP51389 (a growth hormone secretagogue) to assess pituitary responsiveness. No consistent effects (P > 0.1) of treatment were observed for plasma LH, FSH, prolactin, or thyroxine concentrations in samples collected every 3 d. The only effect on ovarian follicle numbers was a reduction in number of follicles 11 to 19 mm in diameter due to TRH treatment (P = 0.029). No mare ovulated during treatment. On the days of frequent sampling, mean LH (P = 0.0001) and FSH (P = 0.001) concentrations were higher in mares receiving GnRH-A and tended to increase from d 1 through 7. In contrast, mean prolactin (P = 0.001) and thyroid-stimulating hormone (P = 0.0001) concentrations were high in mares receiving TRH on d 1 but rapidly decreased thereafter. When mares were administered the secretagogue mixture on d 29, the LH response was greater (P = 0.0002) in mares that had previously received GnRH-A but the FSH response was not affected (P > 0.1); the prolactin response was greater (P = 0.014) and the TSH response was smaller (P = 0.0005) in mares that had previously received TRH. Surprisingly, an immediate growth hormone response to EP51389 was absent ill all mares. In conclusion, daily GnRH-A treatment stimulated plasma LH and FSH concentrations immediately after injection; although no long-term elevation in preinjection concentrations was achieved, the responses gradually increased over time, indicating a stimulation of gonadotropin production and storage. Daily treatment with TRH stimulated plasma TSH and prolactin concentrations, but the response diminished rapidly and was minimal within a few days, indicating a depletion of pituitary stores and little or no stimulation of production. There was no beneficial effect of adding TRH treatment to the daily GnRH-A regimen.
The aim of the present study was two-fold. First, to characterize the secretory profiles of estradiol-17beta and progesterone in relation to the structural changes observed by ultrasonography during follicular dynamics in non-ovulating llamas. Second, to evaluate the effect of exogenous progesterone on follicular activity, in terms of follicle development and hormone production. In experiment one, six adult non-pregnant, non-lactating llamas were examined daily by rectal palpation and transrectal ultrasonography during 70 days. On day 54, intravaginal devices containing 0.33 g of progesterone (CIDRR) were inserted and left in the vagina during 70 days. On day 54, intravaginal devices containing 0.33 g of progesterone (CIDRR) were inserted and left in the vagina during 16 days. The mean duration of a follicular wave was 22.6 +/- 2.5 days. The follicular growth phase (follicles growing from 3 mm to maximum size) averaged 9.2 +/- 2.8 days, the mature phase (follicles around maximum size) 5.2 +/- 1.4 days and regression phase (follicles with decreasing size) 8.2 +/- 2.2 days. Estradiol-17beta plasma concentrations exhibited a similar wave pattern (P < 0.05). In addition, estradiol-17beta peak plasma concentrations (46.9 +/- 3.3 pmol l-1) were attained approximately 12 days after the beginning of the growing phase in connection with maximum follicle size (11.8 +/- 1.6 mm). After CIDR insertion, a rapid increase in plasma progesterone concentrations was observed, with peak concentrations attained on day 1 after insertion. Thereafter, concentrations decreased gradually. Mean follicle size steadily decreased from the day of CIDR insertion to day 11 post-insertion (10.3 +/- 1.6 and 3.3 +/- 0.8 mm, respectively).

In order to investigate the effect of follicle size at CIDR insertion on the outcome of progesterone treatment, experiment two was designed. Sixteen adult non-pregnant and non-lactating llamas were divided into four groups according to follicle development at the time of CIDR insertion (group I: follicles less than or equal to 6 mm; group II: follicles between 6 and 9 mm; group III: follicles between 10 and 14 mm and group IV, regressing follicles). In groups II, III and IV, a significant decrease in follicle size was observed after insertion of the CIDR device. In group I, no further development of dominant follicles was observed until the device was withdrawn. In all cases, the smallest diameter was registered between days 5 and 7 after the beginning of treatment. In conclusion, a detailed characterization of follicular waves using ultrasound and hormone determinations simultaneously in non-ovulating llamas and after the insertion of progesterone releasing devices, is presented.

The Society for Theriogenology does not take responsibility for information contained in or accuracy of the Abstracts published in this newsletter.

Minimum Number of Spermatozoa Required for Normal Fertility After Deep Intrauterine Insemination in Non-sedated Sows

A fibreoptic endoscope procedure for non-surgical deep intrauterine insemination in non-sedated sows has been reported. However, the endoscope is an expensive and fragile instrument, and is unsuitable for use under field conditions. The aim of this study was to determine the minimum number of spermatozoa required to maintain optimal fertility using a flexible catheter (1.8 m in length, 4 mm in diameter) for deep intrauterine insemination in 2-6 parity non-sedated sows. Crossbred sows were treated with eCG 24 h after weaning and with hCG 72 h later to induce estrus. Deep intrauterine insemination was performed 36 h after hCG treatment in 117, 126, 60 and 69 sows with 15.0, 5.0, 2.5 or 1.0 x 107 spermatozoa in 10 ml, respectively. Weaned sows (n=147) not treated with hormones and used for standard artificial insemination (AI) (two inseminations per estrus with 3 x 109 spermatozoa in 100 ml) served as controls. The flexible catheter was passed successfully through the cervix into one uterine horn in 95.4% of the sows in an average of 3.7 +/- 0.09 min. Farrowing rates after deep intrauterine insemination with 15 or 5 x 107 spermatozoa did not differ from those of the control group (82.9, 76.2 and 83.0%, respectively), but a significant decrease (P<0.001) was observed in sows inseminated with 2.5 or 1.0 x 107 spermatozoa (46.7 and 39.1%, respectively). In contrast, the number of spermatozoa inseminated did not affect prolificacy. Laparotomy revealed that the tip of the flexible catheter reached approximately the anterior third of the uterine horn. Although deep intrauterine insemination was performed in only one uterine horn, the percentages of embryos collected from the tip of both uterine horns 2 days after deep insemination were not significantly different. The results show that in comparison with standard AI, a 20-60-fold reduction in the number of spermatozoa inseminated and an 8-10-fold reduction in the dose volume can be achieved without decreasing fertility when semen is deposited non-surgically into the upper first third of one uterine horn.
South American camelids are dribble ejaculators, and urethral contractions occur throughout copulation, which may last 25 min. The urethral contractions and their association with semen characteristics during copulation were determined in llamas and alpacas. A transrectal probe was held in the rectum of the male while copulating an artificial vagina, which was accessed underneath the dummy through a hole. The semen-collecting tube was changed every 5 min. Semen characteristics, color, volume, consistency, motility, concentration, and percentage of live sperm were determined at 5-min intervals. Urethral contractions were evenly distributed during copulation: 40 in alpacas and 63 in llamas (p < .05), with a general range of 11 to 132. semen color was milky in 63% and translucent in 36.5% for alpacas; and creamy (9.9%), milky (47%), and translucent (42%) for llamas. The mean volume of ejaculate was 0.3, 0.4, 0.6, 0.7, 0.6, 0.8, 0.3, and 3.0 mL for 5, 10, 15, 20, 25, and 30 min, respectively. semen consistency was variable: viscous (65%) and semiviscous (34%) in alpacas; and viscous (57%) and semiviscous (42%) in llamas. Spermatic motility varied between 60 and 80% for the llama, and 40 and 80% for the alpaca. Spermatic concentration varied between 60 and 188 x 10^3/mm^3 in llamas, and 30 and 170 x 10^3/mm^3 in alpacas. Percentage of live sperm varied the least: 81 to 90%, in llamas and 65 to 90% in alpacas. The ejaculate of llamas and alpacas is not fractionated, urethral contractions are evenly distributed during copulation, and semen characteristics are present throughout the copulatory period.

In the present study the concentration of relaxin in peripheral blood plasma was assessed during canine pregnancy for its suitability as a pregnancy indicator, using a newly developed relaxin enzyme immunoassay. A significant relaxin increase was found in pregnancy at day 24 after ovulation. However, this relaxin increase did not correlate either with litter size or with body weight of the bitch. Induction of abortion with prostaglandin F-2alpha resulted in reduced peripheral relaxin levels, suggesting a damage of the placenta due to this medical intervention. Thus, the results confirm that relaxin, which is produced by the placenta, is a useful marker for early pregnancy diagnosis in the bitch. Relaxin measurement is recommended for detection of pregnancy either alone, or as supplement of ultrasonographic findings.

To clarify whether red foxes (Vulpes vulpes) can be final hosts of Neospora caninum, foxes and dogs were fed in parallel on tissues of a sheep and a goat experimentally infected with N. caninum. The feces of at least two of five dogs contained N. caninum oocysts, as determined by bioassay. In the feces of all six foxes fed in parallel, oocysts were detected that were larger in size (length 12.6+/-0.5 _m, width 11.8+/-0.4 _m) than the oocysts shed by the dogs. Ribosomal RNA sequences and the results of an immunoblot-based bioassay provided further evidence that these oocysts were different from N. caninum. A titration experiment performed to determine the sensitivity of a bioassay utilizing gerbils showed that as few as five sporulated N. caninum oocysts could be detected by this test. This indicates that, in two feeding experiments, less than 3,700 and 200 sporulated N. caninum oocysts, respectively, could have been among the Hammondia sp.-like oocysts collected from fox feces. These results suggest that the red fox is either an inappropriate final host for N. caninum or not at all a final host for this parasite.
In cattle, the first postpartum dominant follicle has a predilection for the ovary contralateral to the previously gravid uterine horn, possibly due to a local inhibitory effect of the regressing corpus luteum of pregnancy in the ipsilateral ovary. The aim of the present study was to test the hypothesis that the regressing corpus luteum of pregnancy suppresses folliculogenesis in the ipsilateral ovary after parturition. Dairy cows were treated with prostaglandin F₂α between 190 and 220 days of gestation to cause luteolysis without inducing parturition (n = 14) or were untreated controls (n = 32). Follicular growth and function were monitored by daily transrectal ultrasoundography and collection of plasma samples for estimation of FSH, estradiol, and progesterone concentrations. The proportion of first dominant follicles in the ipsilateral ovary was similar for treated and control animals (4/14 vs. 8/32), as was the time interval between calving and establishment of a dominant follicle (mean +/- SEM, 10.1 +/- 0.4 vs. 10.7 +/- 0.5 days). Furthermore, no significant effect of treatment on dominant follicle growth or function was found as determined by plasma hormone concentrations. Although greater folliculogenesis was found in the ovary contralateral to the previously gravid uterine horn, once the location of the future first dominant follicle was selected, the timing of events was independent of location. We suggest that the corpus luteum of pregnancy does not have a local effect on postpartum ovarian folliculogenesis and that, instead, an effect of the previously gravid uterine horn shortly after parturition should be considered.
This study was carried out to investigate the effects of season influencing semen characteristics, frozen-thawed sperm viability and testosterone concentration in Duroc boars. There were no significant differences in the semen volume and sperm concentration of Duroc boars among spring, summer, autumn and winter. However, the pH of sperm-rich and sperm-poor fractions in autumn and winter season was higher than in spring and summer season in Duroc boars. Sperm motility and normal acrosome of raw semen in Duroc boars did not differ significantly among spring, summer, autumn and winter. However, motility and normal acrosome of frozen-thawed sperm were higher in spring season than in summer, autumn and winter. Serum testosterone concentrations in Durocs were higher in spring than summer, autumn and winter. In conclusion, when serum testosterone concentrations were higher, frozen-thawed sperm viability in Duroc boars were higher.


The findings of a retrospective survey of 1393 Thoroughbred mares visiting 22 studfarms in the Newmarket region of the UK during the 1998 mating season were compared with those of a similar study undertaken in 1983. The effects of mare age and status, stallion, month of mating, application of uterine treatments and other parameters on the rates of singleton and twin conception and subsequent pregnancy losses were analyzed. Mare age and status significantly affected the per cycle pregnancy rate and the incidence of pregnancy loss. Overall, the mean number of matings per estrus was 1.12 and the mean number of times a mare was mated until diagnosed pregnant at 15 days after ovulation was 1.88. An overall mean per cycle pregnancy rate of 59.9% at 15 days after ovulation resulted in 94.8% of the mated mares being pregnant at least once at 15 days after ovulation. This high initial pregnancy rate fell to 89.7% by Day 35 and 87.5% by the time of the October pregnancy test; 82.7% of the mares surveyed gave birth to a live foal at term, which compares favorably with the proportion of mares foaling in 1983 (77%). However, despite improvements in the foaling rates over the last 15 years, the overall rate of pregnancy failure remains high and represents a major loss to the Thoroughbred breeding industry.

Reproductive Efficiency of Intensively Managed Thoroughbred Mares in Newmarket MORRIS, LHA; ALLEN, WR (2002): EQUINE. VET. J. 34, 51-60.

Coxiella burnetii infection in pregnant sheep typically causes abortion or the birth of weak lambs. Two C. burnetii-related abortions in a group of 34 pregnant ewes were reported at their first lambing in our research institute. The seroprevalence of C. burnetii infection and bacteria shedding were investigated using an ELISA and PCR, respectively, during the course of two subsequent pregnancies. None of the ewes examined seroconverted from negative to positive at the time of the second and the third parturition and most of the ewes that were seropositive at the abortion episode remained positive throughout the investigation. The two successive pregnancies resulted in the birth of healthy lambs without PCR evidence of coxiella infection from placenta and vaginal swabs taken postpartum. PCR assay performed on vaginal swabs taken from all animals 1, 5 or 12 days after the second lambing were also negative for coxiella. However, one ewe that had previously experienced C. burnetii shedding at the first lambing excreted the bacteria in the genital tract after the third parturition. The bacteria could not be detected by PCR in milk and fecal samples taken up to 12 days after both parturitions.

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